

New Lignans from the Roots of *Taiwania cryptomerioides* HAYATA

by Chiou-Fung Chyu^{a)} and Yueh-Hsiung Kuo^{*b)c)d)}

^{a)} Department of Chemistry, National Taiwan University, Taipei 106, Taiwan

^{b)} Research Center of Food and Biomolecules, National Taiwan University, Taipei 106, Taiwan
(phone: + 886-233661671; fax: + 886-2-23636359; e-mail: yhkue@ntu.edu.tw)

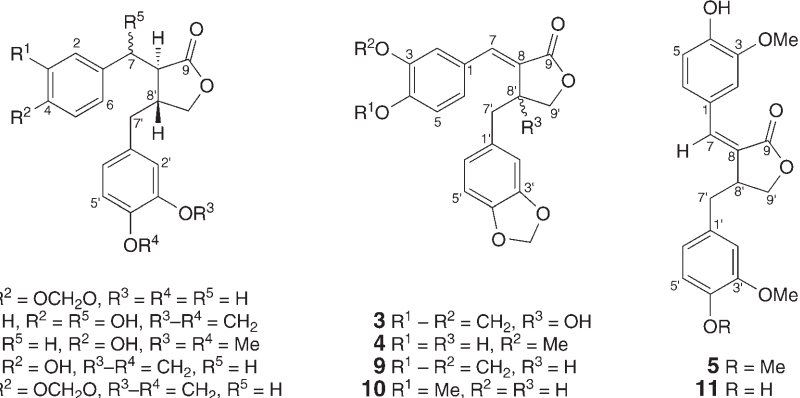
^{c)} College of Pharmacy, China Medical University, Taichung, 404, Taiwan

^{d)} Agriculture Biotechnology Research Center, Academia Sinica, Taipei, 115, Taiwan

The five new lignans designated 3',4'-de-*O*-methylehinokinin (**1**), taiwaninolide (**2**), 8'-hydroxysavinin (**3**), isogumarol (**4**), and 4'-*O*-methyalsalicifolin (**5**), as well as the new 4-(3,4-dimethoxybenzyl)dihydro-3-(4-hydroxybenzyl)furan-2(3*H*)-one (**6**) were isolated from the roots of *Taiwania cryptomerioides*, besides the three known compounds hinokinin (**8**), savinin (**9**), and 3,4-de-*O*-methylehinokinin (**7**). The structures of the new constituents were elucidated through chemical and spectral studies. A compound previously isolated from the heartwood of *Chamaecyparis obtusa* var. *formosana* was assigned structure **1**; however, this structure has now been revised to be 3,4-de-*O*-methylehinokinin (**7**).

Introduction. – *Taiwania cryptomerioides* (Taxodiaceae) is taxonomically included in one genus and one species of endemic plants in Taiwan. It contains more than 6% of essential oil in its heartwood [1]. *T. cryptomerioides* is an important building material with high value in Taiwan. Previously, we investigated the chemical components of the heartwood [2–4] and bark [5–9] of this plant because of its antifungal and decay-resistant characteristics as well as of its beautiful yellowish-red color with distinct purplish-pink streaks. α -Cadinol was found as a major component in its essential oil, which showed selectivity for human-colon-tumor cell lines [10]. Also, it has been a potent agent against wood-decay fungi [11]. Besides, we have found lignans and cadinane-type compounds in this essential oil exhibiting significant cytotoxicity against three human-tumor cell lines [12]. Their interesting structures and those conferring biological activities inspired us to study the chemical components of the roots of *T. cryptomerioides*. Several sesquiterpenes with unique structures, including novel secocadinane-type [13], seco-abeoguaiane-type [14], seco-norabietane-type [15], and norabietane-type [16] derivatives, were isolated from these roots and their structures elucidated. In this paper, we would like to report the five new lignans 3',4'-de-*O*-methylehinokinin (**1**), taiwaninolide (**2**), 8'-hydroxysavinin (**3**), isogumarol (**4**), and 4'-*O*-methyalsalicifolin (**5**), and the new 4-(3,4-dimethoxybenzyl)dihydro-3-(4-hydroxybenzyl)furan-2(3*H*)-one (**6**), together with the three known lignans 3,4-de-*O*-methylehinokinin (**7**) [17], hinokinin (**8**) [18], and savinin (**9**) [19]¹⁾. One compound, isolated from the heartwood of *Chamaecyparis obtusa* var. *formosana*,

¹⁾ Trivial names and trivial atom numbering; for systematic names, see *Exper. Part*.



was previously assigned the structure **1**; this assignment has now been revised to the structure of 3,4-de-*O*-methylenehinokinin (**7**).

Results and Discussion. – Compound **1** was isolated as a pale yellow gum, which had a molecular formula $\text{C}_{19}\text{H}_{18}\text{O}_6$, as established by analysis of its ^{13}C -NMR spectrum (Table 1) and HR-EI-MS. The IR spectrum of **1** confirmed the presence of an OH group (3410 cm^{-1}), a γ -lactone carbonyl group (1745 cm^{-1}), and aromatic moieties ($1608, 1489\text{ cm}^{-1}$). Detailed analysis of the NMR (Table 1), MS (Fig. 1), HMBC, and NOESY (Fig. 2) data established the structure of **1** as (8*R*,8'*R*)-3',4'-de-*O*-methylenehinokinin¹. Comparison of the ^1H - and ^{13}C -NMR data of **1** and hinokinin (**8**) also verified the structure. The $\text{CH}_2(9')$ protons were nonequivalent [18], as two benzyl groups were in *trans*-configuration. Therefore, compound **1** was in *trans*-configuration. The value of the specific rotation of **1** ($[\alpha]_{\text{D}} = 20.2$) was near that of (–)-hinokinin ($[\alpha]_{\text{D}} = -26.3$) which confirmed the (8*R*,8'*R*) configuration of **1**, and the CD curve of **1** ($[\Delta\epsilon]_{234} - 2.12$, $[\Delta\epsilon]_{286} - 0.25$) was closely similar to the ORD curve of (8*R*,8'*R*)-(–)-hinokinin ($[\phi]_{250} - 10900$, $[\phi]_{285} + 400$, $[\phi]_{300} - 4850$) [18][20].

The ^1H -NMR spectrum of **1** (Table 1) exhibited signals for a methylenedioxy group ($\delta 5.92$ (*d*, $J = 1.2\text{ Hz}$, 1 H) and 5.91 (*d*, $J = 1.2\text{ Hz}$, 1 H)) at a benzene moiety and CH_2 protons in γ -position of a γ -lactone ring ($\delta 3.84$ (*dd*, $J = 9.2, 7.2\text{ Hz}$, 1 H) and 4.09 (*dd*, $J = 9.2, 7.2\text{ Hz}$, 1 H)). Also, two *ABX* systems (6 H) of aromatic protons were observed ($\delta 6.59$ (*d*, $J = 1.6\text{ Hz}$, 1 H), 6.71 (*d*, $J = 8.0\text{ Hz}$, 1 H), and 6.57 (*dd*, $J = 8.0, 1.6\text{ Hz}$, 1 H); 6.50 (*d*, $J = 2.0\text{ Hz}$, 1 H), 6.74 (*d*, $J = 8.0\text{ Hz}$, 1 H), and 6.42 (*dd*, $J = 8.0, 2.0\text{ Hz}$, 1 H)), besides four benzylic protons ($\delta 2.82$ (*dd*, $J = 14.0, 7.2\text{ Hz}$, 1 H), 2.93 (*dd*, $J = 14.0, 4.8\text{ Hz}$, 1 H); $2.38-2.45$ (*m*, 1 H), $2.51-2.55$ (*m*, 1 H)). The ^{13}C -NMR spectrum and DEPT experiment showed 19 signals including a γ -lactone carbonyl C-atom ($\delta 179.0$), twelve aromatic C-atoms, and four CH_2 ($\delta 101.0, 71.4, 37.9, 34.7$) and two CH groups ($\delta 46.5, 41.1$). The MS of **1** displayed a base peak at m/z 135 and a peak at m/z 219 (see Fig. 1), in accordance with a fragment ion of an 8-[(3,4-methylenedioxy)benzyl]-substituted lactone moiety. Another peak at m/z 123 corresponded to a 3,4-dihydroxybenzyl cation. The signals at $\delta 2.82$ and 2.93 were assigned to $\text{CH}_2(7)$, due to a lower-field location than those of $\text{CH}_2(7')$ and to the HMBC correlations to the lactone carbonyl group ($\delta 179.0$, C(9)), C(1), C(2), and C(6). The $\text{CH}_2(7')$ also have HMBC correlations to C(1'), C(2'), C(6'), and C(9'). In the HMBC plot, the OCH_2O protons correlated to C(3) and C(4), and the NOESY correlations (Fig. 2) (H–C(7)/H–C(2), H–C(6); H–C(7')/H–C(2'), H–C(6')) confirmed the structure of **1** as shown.

Table 1. ^1H - and ^{13}C -NMR Data (CDCl_3 , 400 and 100 MHz) of Compounds **1**–**3**¹). δ in ppm, J in Hz.

	1		2		3	
	$\delta(\text{H})$	$\delta(\text{C})$	$\delta(\text{H})$	$\delta(\text{C})$	$\delta(\text{H})$	$\delta(\text{C})$
C(1)		131.3 (s)		132.8 (s)		127.1 (s)
H–C(2)	6.59 (<i>d</i> , $J = 1.6$)	109.5 (<i>d</i>)	7.12 (<i>d</i> , $J = 8.8$)	126.6 (<i>d</i>)	7.67 (<i>d</i> , $J = 1.6$)	111.3 (<i>d</i>)
C(3) or H–C(3)		147.8 (s)	6.77 (<i>d</i> , $J = 8.8$)	115.4 (s)		148.2 (s)
C(4)		146.4 (s)		155.3 (s)		149.9 (s)
H–C(5)	6.71 (<i>d</i> , $J = 8.0$)	108.3 (<i>d</i>)	6.77 (<i>d</i> , $J = 8.8$)	115.4 (<i>d</i>)	6.84 (<i>d</i> , $J = 8.0$)	108.5 (<i>d</i>)
H–C(6)	6.57 (<i>dd</i> , $J = 8.0, 1.6$)	122.3 (<i>d</i>)	7.12 (<i>d</i> , $J = 8.8$)	126.6 (<i>d</i>)	7.30 (<i>dd</i> , $J = 8.0, 1.6$)	128.9 (<i>d</i>)
CH ₂ (7) or H–C(7)	2.82 (<i>dd</i> , $J = 14.0, 7.2$), 2.93 (<i>dd</i> , $J = 14.0, 4.8$)	34.7 (<i>t</i>)	5.27 (<i>d</i> , $J = 3.2$)	71.7 (<i>d</i>)	7.63 (s)	143.0 (<i>d</i>)
H–C(8) or C(8)	2.47–2.53 (<i>m</i>)	46.5 (<i>d</i>)	2.60 (<i>dd</i> , $J = 6.8, 3.2$)	52.7 (<i>d</i>)		126.2 (s)
C(9)		179.0 (s)		178.5 (s)		171.2 (s)
C(1')		130.7 (s)		131.5 (s)		128.1 (s)
H–C(2')	6.50 (<i>d</i> , $J = 2.0$)	115.6 (<i>d</i>)	6.26 (<i>d</i> , $J = 1.6$)	108.6 (<i>d</i>)	6.68 (<i>d</i> , $J = 1.6$)	110.7 (<i>d</i>)
C(3')		143.9 (s)		147.7 (s)		147.9 (s)
C(4')		142.4 (s)		146.2 (s)		147.1 (s)
H–C(5')	6.74 (<i>d</i> , $J = 8.0$)	115.4 (<i>d</i>)	6.60 (<i>d</i> , $J = 8.0$)	108.1 (<i>d</i>)	6.69 (<i>d</i> , $J = 8.0$)	108.4 (<i>d</i>)
H–C(6')	6.42 (<i>dd</i> , $J = 8.0, 2.0$)	120.9 (<i>d</i>)	6.32 (<i>dd</i> , $J = 8.0, 1.6$)	121.5 (<i>d</i>)	6.62 (<i>dd</i> , $J = 8.0, 1.6$)	123.6 (<i>d</i>)
CH ₂ (7')	2.38–2.45 (<i>m</i>), 2.51–2.55 (<i>m</i>)	37.9 (<i>t</i>)	2.23 (<i>dd</i> , $J = 14.0, 7.2$), 2.34 (<i>dd</i> , $J = 14.0, 8.4$)	39.2 (<i>t</i>)	2.97 (<i>d</i> , $J = 14.0$), 3.22 (<i>d</i> , $J = 14.0$)	41.8 (<i>t</i>)
H–C(8') or C(8')	2.41–2.47 (<i>m</i>)	41.1 (<i>d</i>)	2.72–2.81 (<i>m</i>)	36.4 (<i>d</i>)		76.5 (s)
CH ₂ (9')	3.84 (<i>dd</i> , $J = 9.2, 7.2$), 4.09 (<i>dd</i> , $J = 9.2, 7.2$)	71.4 (<i>t</i>)	3.90 (<i>dd</i> , $J = 8.8, 6.4$), 4.26 (<i>dd</i> , $J = 8.8, 8.0$)	72.5 (<i>t</i>)	3.95 (<i>d</i> , $J = 9.6$), 4.45 (<i>d</i> , $J = 9.6$)	76.0 (<i>t</i>)
OCH ₂ O	5.91 (<i>d</i> , $J = 1.2$), 5.92 (<i>d</i> , $J = 1.2$)	101.0 (<i>t</i>)			6.01 (s)	101.8 (<i>t</i>)
OCH ₂ O			5.88 (<i>d</i> , $J = 1.6$), 5.91 (<i>d</i> , $J = 1.6$)	101.0 (<i>t</i>)	5.91 (<i>d</i> , $J = 1.6$), 5.92 (<i>d</i> , $J = 1.6$)	101.1 (<i>t</i>)
OH	5.52, 6.71 (2 br. s)					

Taiwaninolide (**2**) had a molecular ion at m/z 342.1108 in the HR-EI-MS, as analyzed for $\text{C}_{19}\text{H}_{18}\text{O}_6$. Analysis of its IR spectrum suggested that **2** contains an OH group (3410 cm^{-1}), a γ -lactone carbonyl group (1751 cm^{-1}), and an aromatic moiety ($1615, 1492\text{ cm}^{-1}$). Eleven indices of hydrogen deficiency (IHD) were determined from the molecular formula, the ^{13}C -NMR spectrum (Table 1), and the DEPT experiment. Further spectral data (Table 1, Figs. 1 and 2) established the structure of **2** as *trans*-4-(1,3-benzodioxol-5-ylmethyl)dihydro-3-[hydroxy(4-hydroxyphenyl)methyl]-furan-2(3*H*)-one.

The ^1H -NMR spectrum (Table 1) of **2** indicated the presence of an *ABX* system (δ 6.32 (*dd*, $J = 8.0, 1.6\text{ Hz}$, 1 H), 6.60 (*d*, $J = 8.0\text{ Hz}$, 1 H), and 6.26 (*d*, $J = 1.6\text{ Hz}$, 1 H)), a symmetrical A_2X_2 pattern for aromatic protons (δ 7.12 (*d*, $J = 8.8\text{ Hz}$, 2 H) and 6.77 (*d*, $J = 8.8\text{ Hz}$)), and a OCH₂O group (δ 5.88 (*d*,

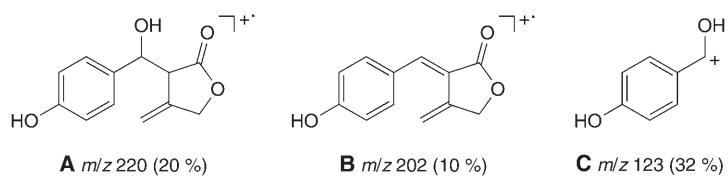
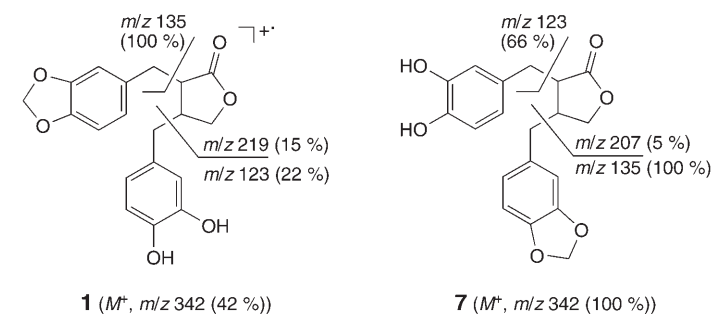


Fig. 1. MS Fragments

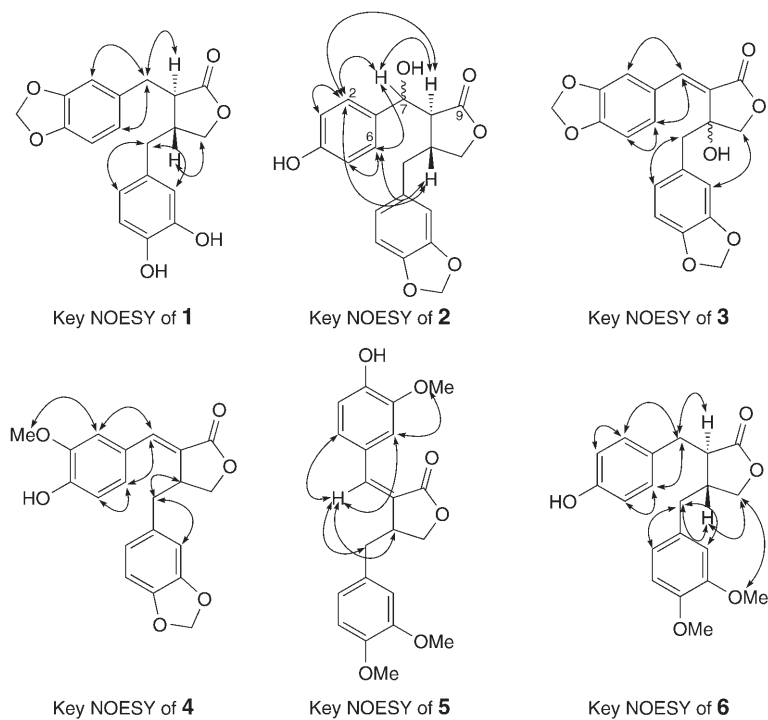


Fig. 2. Key NOESY correlations of 1–6

$J = 1.6$ Hz, 1 H) and 5.91 ($d, J = 1.6$ Hz, 1 H)). The ^{13}C -NMR and DEPT established the presence of a γ -lactone carbonyl group (δ 178.5), two aromatic rings, and an sp^3 -oxygenated C-atom (δ 71.7). The latter and a proton at δ 5.27 ($d, J = 3.2$ Hz) were assigned to the benzylic position CH(7). Two CH_2 groups (δ 3.90 ($dd, J = 8.8, 6.4$ Hz) and 4.26 ($dd, J = 8.8, 8.0$ Hz); δ 2.23 ($dd, J = 14.0, 7.2$ Hz) and 2.34 ($dd, J = 14.0, 8.4$ Hz)) are characteristic of a CH_2 group of a γ -lactone and of a benzyl group, respectively. From the molecular formula and the spectral evidence, compound **2** was consistent with a dibenzyl-substituted γ -butyrolactone. The base peak in the MS at m/z 135 and three peaks at m/z 220, 202, and 123 (see fragment ions **A–C** in Fig. 1) revealed the presence of a 3,4-(methylenedioxy)benzyl moiety at C(8') and of a 4,7-dihydroxybenzyl moiety at C(8). The HMBC correlations H–C(7)/C(6), C(2), and C(9), H–C(7')/C(8), C(8'), C(9'), C(2'), and C(6') confirmed the gross structure. The NOESY correlations (Fig. 2), together with the nonequivalent protons $\text{CH}_2(9')$, suggested that the two benzyl moieties were in *trans*-configuration [18].

The 8'-hydroxysavinin (**3**) was isolated as a yellowish gum; its molecular formula $\text{C}_{20}\text{H}_{16}\text{O}_7$ was established by ^{13}C -NMR and HR-EI-MS data, and the IHD was 13. The IR spectrum of **3** showed absorptions attributable to an OH group (3443 cm^{-1}), to a γ -lactone carbonyl group (1747 cm^{-1}), and to aromatic moieties ($1628, 1497\text{ cm}^{-1}$). The ^{13}C - and ^1H -NMR spectrum (Table 1) data together with the UV absorption bands at λ_{max} 239, 258, 290, and 336 nm suggested that compound **3** is very similar to savinin (**9**) [19]. Based on further spectral data, the structure of **3** was established to be 8'-hydroxysavinin.

The following HMBC correlations were observed for **3**: H–C(7)/C(2), C(6), C(9), and C(8'); $\text{CH}_2(7')$ /C(2'), C(6'), and C(8'), and H–C(6)/C(4), C(5), and C(7). The NOESY correlations (Fig. 2) H–C(7)/H–C(2) and H–C(6), H–C(5)/H–C(6), H–C(6')/H–C(7'), and H–C(9')/H–C(2') confirmed the assigned structure and relative configuration. Thus, the difference between **3** and **9** was that **3** has an additional OH group at C(8'). This caused a shift to lower field of the $\text{CH}_2(7')$ and $\text{CH}_2(9')$ signals as compared to those of **9**, accompanied by a simplification of their pattern to two *AX* systems (δ 2.97, 3.22 ($2d, J = 14.0$ Hz, 1 H each, $\text{CH}_2(7')$) and 3.95, 4.45 ($2d, J = 9.6$ Hz, 1 H each, $\text{CH}_2(9')$)) and a low-field shift of the C(7) and C(9) signals as compared to those of **9**.

Based on the HR-EI-MS and ^{13}C -NMR data (Table 2), isoguamarol (**4**) has the molecular formula $\text{C}_{20}\text{H}_{18}\text{O}_6$, with an IHD of 12. The IR spectrum of **4** displayed peaks for an OH group (3423 cm^{-1}), a γ -lactone carbonyl group (1744 cm^{-1}), and aromatic moieties (1595 and 1518 cm^{-1}). The UV absorption bands at λ_{max} 237, 296, and 332 nm, the ^{13}C -NMR and DEPT data (Table 2), in addition to the base peak in the MS at m/z 135 (3,4-methylenedioxybenzyl cation) indicated that compound **4** is a positional isomer of guamarol (**10**) [21].

The ^1H -NMR signals (Table 2) of **4** indicated the presence of two *ABX* systems of aromatic protons, an OCH_2O group at a benzene moiety, a proton at a trisubstituted olefin moiety (δ 7.49 ($d, J = 1.6$ Hz, 1 H)), an MeO group, and a phenolic proton (δ 6.02 (br. s, 1 H; D_2O exchangeable)). The HMBC correlations H–C(7)/C(9), C(2), and C(6), and $\text{CH}_2(7')$ /C(8'), C(2'), and C(6') confirmed the assigned structure. Based on the NOESY correlations (Fig. 2), the MeO group showed a NOESY correlation with H–C(2), which established the position of the MeO group at C(3).

Compound **5** has the formula $\text{C}_{21}\text{H}_{22}\text{O}_6$ according to the HR-EI-MS and ^{13}C -NMR data. It has an IHD of 11 as deduced from its molecular formula. The IR spectrum shows absorptions for an OH group (3418 cm^{-1}), a conjugated γ -lactone carbonyl group (1740 cm^{-1}), and an aromatic moiety (1595 and 1517 cm^{-1}). The UV spectra of **5**

Table 2. ^1H - and ^{13}C -NMR Data (CDCl_3 , 400 and 100 MHz) of Compounds **4**–**6**^a. δ in ppm, J in Hz.

	4		5		6	
	$\delta(\text{H})$	$\delta(\text{C})$	$\delta(\text{H})$	$\delta(\text{C})$	$\delta(\text{H})$	$\delta(\text{C})$
C(1)		126.4 (s)		126.5 (s)		130.5 (s)
H–C(2)	7.00 (<i>d</i> , $J=2.0$)	112.6 (<i>d</i>)	8.21 (<i>d</i> , $J=1.6$)	113.3 (<i>d</i>)	6.96 (<i>d</i> , $J=8.4$)	130.4 (<i>d</i>)
C(3) or H–C(3)		146.5 (s)		146.1 (s)	6.72 (<i>d</i> , $J=8.4$)	115.5 (<i>d</i>)
C(4)		146.7 (s)		147.7 (s)		154.6 (s)
H–C(5)	6.98 (<i>d</i> , $J=8.0$)	115.0 (<i>d</i>)	6.84 (<i>d</i> , $J=8.4$)	113.9 (<i>d</i>)	6.72 (<i>d</i> , $J=8.4$)	115.5 (<i>d</i>)
H–C(6)	7.17 (<i>dd</i> , $J=8.0, 1.6$)	123.8 (<i>d</i>)	6.98 (<i>dd</i> , $J=8.4, 2.0$)	126.6 (<i>d</i>)	6.96 (<i>d</i> , $J=8.4$)	130.4 (<i>d</i>)
H–C(7) or CH ₂ (7)	7.49 (<i>d</i> , $J=1.6$)	137.6 (<i>d</i>)	6.54 (<i>d</i> , $J=1.6$)	141.0 (<i>d</i>)	2.85 (<i>dd</i> , $J=14.2, 7.2$), 2.95 (<i>dd</i> , $J=14.2, 5.6$)	34.1 (<i>t</i>)
C(8) or H–C(8)		125.3 (s)		124.3 (s)	2.53–2.56 (<i>m</i>)	46.5 (<i>d</i>)
C(9)		172.7 (s)		169.7 (s)		178.7 (s)
C(1')		131.5 (s)		130.4 (s)		129.7 (s)
H–C(2')	6.63 (<i>d</i> , $J=1.6$)	109.0 (<i>d</i>)	6.67 (<i>d</i> , $J=2.0$)	112.9 (<i>d</i>)	6.43 (<i>d</i> , $J=1.6$)	111.7 (<i>d</i>)
C(3')		148.0 (s)		149.3 (s)		149.0 (s)
C(4')		147.5 (s)		148.2 (s)		147.8 (s)
H–C(5')	6.71 (<i>d</i> , $J=8.0$)	108.5 (<i>d</i>)	6.80 (<i>d</i> , $J=8.0$)	111.8 (<i>d</i>)	6.74 (<i>d</i> , $J=8.0$)	111.3 (<i>d</i>)
H–C(6')	6.61 (<i>dd</i> , $J=8.0, 1.6$)	122.0 (<i>d</i>)	6.71 (<i>dd</i> , $J=8.0, 2.0$)	121.4 (<i>d</i>)	6.54 (<i>dd</i> , $J=8.0, 1.6$)	120.6 (<i>d</i>)
CH ₂ (7')	2.58 (<i>dd</i> , $J=14.4, 10.4$), 3.01 (<i>dd</i> , $J=14.4, 4.4$)	37.4 (<i>t</i>)	2.80 (<i>dd</i> , $J=13.8, 8.6$), 2.91 (<i>dd</i> , $J=13.8, 7.0$)	40.8 (<i>t</i>)	2.44–2.49 (<i>m</i>), 2.57–2.61 (<i>m</i>)	38.2 (<i>t</i>)
H–C(8')	3.75–3.79 (<i>m</i>)	39.7 (<i>d</i>)	3.25–3.31 (<i>m</i>)	44.5 (<i>d</i>)	2.41–2.47 (<i>m</i>)	41.1 (<i>d</i>)
CH ₂ (9')	4.24 (<i>br. d</i> , $J=4.0$)	69.5 (<i>t</i>)	4.10 (<i>dd</i> , $J=9.0, 3.6$), 4.32 (<i>dd</i> , $J=9.0, 7.2$)	70.0 (<i>t</i>)	3.86 (<i>dd</i> , $J=8.8, 7.2$), 4.11 (<i>dd</i> , $J=8.8, 7.2$)	71.3 (<i>t</i>)
OCH ₂ O	5.91 (<i>d</i> , $J=1.2$), 5.92 (<i>d</i> , $J=1.2$)	101.1 (<i>t</i>)				
MeO–C(3)	3.91 (s)	56.0 (<i>q</i>)	3.92 (s)	56.1 (<i>q</i>)		
MeO–C(3')			3.80 (s)	56.0 (<i>q</i>)	3.79 (s)	55.8 (<i>q</i>)
MeO–C(4')			3.85 (s)	56.1 (<i>q</i>)	3.84 (s)	55.9 (<i>q</i>)
OH	6.02 (<i>br. s</i>)		5.95 (s)			

confirmed the presence of a conjugated carbonyl group (λ_{max} 287 and 336 nm). Analysis of the HMQC, HMBC, and NOESY (Fig. 2) data revealed that the structure of **5** was similar to that of salicifoline (**11**) [22], the only difference being an MeO instead of an OH group at C(4'). Therefore, the structure of **5** was elucidated as 4'-O-methylsalicifolin.

The ^1H -NMR data of **5** (Table 2) indicated the presence of a 3,4-dimethoxybenzyl moiety (*ABX* system of aromatic protons), which was confirmed by the presence of the base peak in the MS at m/z

151), of a 1,3,4-trisubstituted benzene moiety carrying an OH and an MeO group, and a γ -lactone with a methylene group in γ -position. These features are compatible with a dibenzylbutyrolactone lignan. The UV absorption showed a C=C bond linking an aromatic and a lactone carbonyl group, but the olefinic proton (H-C(7)) appeared at higher field than the corresponding proton of compounds **3** and **4**. Due to the deshielding effect of the lactone carbonyl group, H-C(2) was shifted downfield to δ 8.21. These two evidences suggested the (*Z*)-configuration at the exocyclic C(7)=C(8) bond.

Compound **6** exhibited the IR absorption of an OH group (3426 cm^{-1}), a γ -lactone carbonyl group (1765 cm^{-1}), and an aromatic moiety (1615 , 1596 , and 1517 cm^{-1}), and an M^+ at m/z 342.1467 establishing the molecular formula $\text{C}_{20}\text{H}_{22}\text{O}_5$. The specific rotation of **6** was near to that of (–)-(8*R*,8'*R*)-hinokinin (**8**) and the CD curve of **6** closely similar to the ORD curve of **8** [18][20], suggesting that compound **6** has the same (8*R*,8'*R*)-configuration¹) as **8**. Thus, the new compound **6** was (3*R*,4'*R*)-4-(3,4-dimethoxybenzyl)dihydro-3-(4-hydroxybenzyl)furan-2(3*H*)-one [23].

The ¹H-NMR spectrum of **6** (Table 2) showed signals of an *ABX* systems of aromatic protons, a symmetrical A_2X_2 pattern of aromatic protons, and two MeO signals. In the MS, the fragment ions at m/z 151 and 107 revealed the presence of a 3,4-dimethoxybenzyl and a 4-hydroxybenzyl moiety. The $\text{CH}_2(9')$ were nonequivalent (δ 3.86 (*dd*, $J = 8.8, 7.2\text{ Hz}$, 1 H) and 4.11 (*dd*, $J = 8.8, 7.2\text{ Hz}$, 1 H) indicating that the two benzyl moieties were in *trans*-relation to each other [18]. HMBC Correlations (H-C(7)/C(9); H-C(7)/C(9')) and NOESY correlations (Fig. 2) (H-C(7)/H-C(2), H-C(6); H-C(7)/H-C(2'), H-C(6')) determined the correct position of the two benzyl moieties.

The assignments of the ¹³C- and ¹H-NMR data (Table 3) of **7** were supported by COSY, HMQC, HMBC, and NOESY data. Compound **7** was assigned as 3,4-de-*O*-

Table 3. ¹H- and ¹³C-NMR Data (CDCl₃, 400 and 100 MHz) of Compound **7**¹). δ in ppm, J in Hz.

7		
	$\delta(\text{H})$	$\delta(\text{C})$
C(1)		130.1 (<i>s</i>)
H-C(2)	6.67 (<i>d</i> , $J = 2.0$)	116.1 (<i>d</i>)
C(3)		143.8 (<i>s</i>)
C(4)		142.8 (<i>s</i>)
H-C(5)	6.77 (<i>d</i> , $J = 8.0$)	115.3 (<i>d</i>)
H-C(6)	6.54 (<i>dd</i> , $J = 8.0, 2.0$)	121.8 (<i>d</i>)
$\text{CH}_2(7)$	2.86 (<i>br. d</i> , $J = 6.0$)	34.1 (<i>t</i>)
H-C(8)	2.50–2.57 (<i>m</i>)	46.5 (<i>d</i>)
C(9)		179.3 (<i>s</i>)
C(1')		131.6 (<i>s</i>)
H-C(2')	6.46 (<i>d</i> , $J = 2.0$)	108.9 (<i>d</i>)
C(3')		147.8 (<i>s</i>)
C(4')		146.3 (<i>s</i>)
H-C(5')	6.68 (<i>d</i> , $J = 8.0$)	108.4 (<i>d</i>)
H-C(6')	6.45 (<i>dd</i> , $J = 8.0, 2.0$)	121.6 (<i>d</i>)
$\text{CH}_2(7')$	2.41–2.47 (<i>m</i>), 2.57–2.64 (<i>m</i>)	38.2 (<i>t</i>)
H-C(8')	2.45–2.52 (<i>m</i>)	41.0 (<i>d</i>)
$\text{CH}_2(9')$	3.83 (<i>dd</i> , $J = 9.2, 7.6$), 4.07 (<i>dd</i> , $J = 9.2, 7.2$)	71.4 (<i>t</i>)
OCH_2O	5.91 (<i>d</i> , $J = 1.2$), 5.92 (<i>d</i> , $J = 1.2$)	101.0 (<i>t</i>)
OH	5.60, 6.06 (2 <i>br. s</i>)	

methylenehinokinin. To a compound formerly isolated from *Chamaecyparis obtusa* var. *formosana* [17], structure **1** was attributed. But this compound and compound **7** have almost identical physical data (including NMR and $[\alpha]_D$). Thus, the structure of the compound isolated from *Chamaecyparis obtusa* must be revised to that of **7**.

The HMBC correlations (Fig. 3) of **7** included those of the methylenedioxy protons to C(3') and C(4'), and the NOESY correlations (Fig. 3) H–C(7)/H–C(2) and H–C(6), and H–C(7)/H–C(2') and H–C(6'') were observed. The MS of **7** displayed peaks at m/z 123 and 207 (see Fig. 1), in accordance with the 3,4-dihydroxybenzyl group at the lactone atom C(8). Another peak at m/z 135 corresponded to the 3,4-(methylenedioxy)benzyl group.

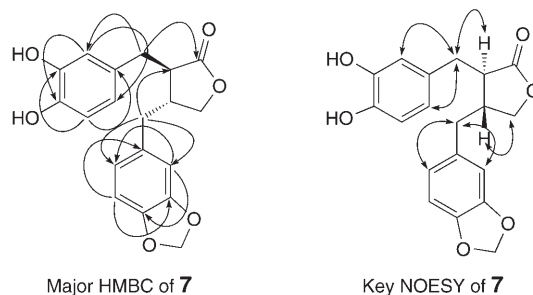


Fig. 3. Major HMBC and key NOESY correlations for **7**

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Experimental Part

General. Column chromatography (CC): silica gel (*Merck*; 70–230 mesh, 230–400 mesh, ASTM). Semi-prep. normal-phase HPLC: column *LiChrosorb Si 60* (250 × 10 mm, 7 μm); *LDC Analytical-III* apparatus. M.p.: *Yanagimoto* micro-melting point apparatus; uncorrected. Specific rotation: *Jasco DIP-180* digital polarimeter. IR Spectra: *Perkin-Elmer 983-G* spectrophotometer; in cm^{-1} . ^1H - and ^{13}C -NMR Spectra: *Varian Unity-400* spectrophotometer. EI-MS: *Jeol JMS-HX-300* mass spectrometer.

Plant Material. The roots of *T. cryptomerioides* were collected from Taichung, Taiwan, in August 1996. The plant was identified by Dr. *Shang-Tzen Chang*, Professor of the Department of Forestry, National Taiwan University. A voucher specimen (no. 013542) has been deposited in the Herbarium of the Department of Botany of the National Taiwan University, Taipei, Taiwan.

Extraction and Isolation. Air-dried root slices of *T. cryptomerioides* (15 kg) were extracted two times with acetone (125 l) at r.t. (7 days twice). The acetone extract was concentrated to give a black residue, which was suspended in H_2O (7 l) and then partitioned (3 times) with 1 l of AcOEt. The AcOEt fraction (365 g) was subjected to CC (silica gel, hexane/AcOEt of increasing polarity). The 8'-hydroxysavinin (**3**; 3 mg), isoguamarol (**4**; 10 mg), 3,4-de-*O*-methylenehinokinin (**7**; 8.0 mg), and hinokinin (**8**; 11.2 mg) were obtained with 30% AcOEt/hexane (CC) and purified by HPLC (10% AcOEt/ CH_2Cl_2 and 20% acetone/hexane). The 3',4'-de-*O*-methylenehinokinin (**1**; 8 mg), taiwaninolide (**2**; 10 mg), 4'-*O*-methylsalicifolin (**5**; 13 mg), and 3-(4-hydroxybenzyl)-4-(3,4-dimethoxybenzyl)dihydrofuran-2-one (**6**; 5 mg) were eluted with 50% AcOEt/hexane (CC) and purified by HPLC (25% AcOEt/ CH_2Cl_2 and 30% acetone/hexane). Savinin (**9**; 2.3 g) was eluted with 20% AcOEt/hexane (CC).

3',4'-De-*O*-methylenehinokinin (= (3*R*,4*R*)-3-(1,3-Benzodioxol-5-ylmethyl)-4-[(3,4-dihydroxyphenyl)methyl]dihydrofuran-2(3*H*)-one; **1**): Pale yellow gum. $[\alpha]_D^{25} = -20.2$ ($c = 0.24$, CHCl_3). UV (MeOH): 230.0 (3.8), 285.0 (3.7). CD (MeOH): -2.12 (234), -0.25 (286). IR (KBr): 3410, 1745,

1608, 1502, 1489, 1284, 1193. ¹H- and ¹³C-NMR (CDCl₃, 400, 100 MHz): Table 1. EI-MS: 342 (42, M⁺), 219 (15), 218 (24), 192 (12), 135 (100), 123 (22). HR-EI-MS: 342.1108 (C₁₉H₁₈O₆⁺; calc. 342.1103).

Taiwaninolide (=trans-4-(1,3-Benzodioxol-5-ylmethyl)dihydro-3-[hydroxy(4-hydroxyphenyl)methyl]furan-2(3H)-one; **2**): Yellow gum. [α]_D²⁵ = -57.2 (c = 0.42, CHCl₃). UV (MeOH): 226.5 (4.01), 284.5 (3.65). IR (KBr): 3410, 1751, 1615, 1492, 1446, 1376, 1246. ¹H- and ¹³C-NMR (CDCl₃, 400, 100 MHz): Table 1. EI-MS: 342 (8, M⁺), 324 (26), 220 (20), 202 (10), 123 (32), 189 (6), 135 (100). HR-EI-MS: 342.1108 (C₁₉H₁₈O₆⁺; calc. 342.1103).

8'-Hydroxysavinin (= (3E)-4-(1,3-Benzodioxol-5-ylmethyl)-3-[(1,3-benzodioxol-5-ylmethylene)dihydro-4-hydroxyfuran-2(3H)-one]; **3**): Yellowish gum. [α]_D²⁷ = -34.9 (c = 0.12, CHCl₃). UV (MeOH): 238.5 (4.08), 258.0 (3.94), 289.5 (3.93), 336.0 (3.77). IR (KBr): 3443, 1747, 1628, 1497, 1450, 1248, 1039. ¹H- and ¹³C-NMR (CDCl₃, 400, 100 MHz): Table 1. EI-MS: 368 (7, M⁺), 350 (100), 348 (69), 233 (75), 135 (72). HR-EI-MS: 368.0879 (C₂₀H₁₆O₇⁺; calc. 368.0895).

Isoquamarol (= (3E,4R)-4-(1,3-Benzodioxol-5-ylmethyl)dihydro-3-[(4-hydroxy-3-methoxyphenyl)methylene]furan-2(3H)-one; **4**): Yellow gum. [α]_D²⁵ = -67.8 (c = 0.40, CHCl₃). UV (MeOH): 237.0 (4.09), 295.5 (4.01), 331.5 (4.16). IR (KBr): 3423, 1744, 1645, 1596, 1518, 1447, 1362. ¹H- and ¹³C-NMR (CDCl₃, 400, 100 MHz): Table 2. EI-MS: 354 (18, M⁺), 219 (78), 135 (100). HR-EI-MS: 354.1113 (C₂₀H₁₈O₆⁺; calc. 354.1103).

4'-O-Methylsalicifolin (= (3Z,4R)-4-[(3,4-Dimethoxyphenyl)methyl]dihydro-3-[(4-hydroxy-3-methoxyphenyl)methylene]furan-2(3H)-one; **5**): Yellow gum. [α]_D²⁵ = -26.4 (c = 1.08, CHCl₃). UV (MeOH): 286.5 (2.92), 336.0 (3.12). IR: 3418, 1740, 1638, 1595, 1517, 1465, 1387. ¹H- and ¹³C-NMR (CDCl₃, 400, 100 MHz): Table 2. EI-MS: 370 (24, M⁺), 219 (20), 151 (100). HR-EI-MS: 370.1414 (C₂₁H₂₂O₆⁺; calc. 370.1417).

(3R,4R)-4-[(3,4-Dimethoxyphenyl)methyl]dihydro-3-[(4-hydroxyphenyl)methyl]furan-2(3H)-one; (**6**): Yellow gum. [α]_D²⁵ = -25.9 (c = 0.20, CHCl₃). UV (MeOH): 226.5 (4.09), 280.0 (3.64). ORD (MeOH, c = 0.05; ϕ (λ in nm): +4560 (209), -3090 (231), +3340 (249). IR (KBr): 3426, 1765, 1615, 1596, 1517, 1455, 1266, 1239. ¹H- and ¹³C-NMR (CDCl₃, 400, 100 MHz): Table 2. EI-MS: 342 (100, M⁺), 151 (60), 137 (40), 107 (40). HR-EI-MS: 342.1467 (C₂₀H₂₂O₅⁺; calc. 342.1468).

3,4-De-O-methylenehinokinin (= (3R,4R)-4-(1,3-Benzodioxol-5-ylmethyl)-3-(3,4-dihydroxyphenyl)dihydrofuran-2(3H)-one; **7**): Pale yellow gum. [α]_D²⁵ = -13.2 (c = 0.32, CHCl₃). UV (MeOH): 285.0 (3.80). ORD (MeOH, c = 0.07; ϕ (λ in nm)): -16460 (204), -5120 (235), +40 (256), -1020 (289). IR (KBr): 3445, 1757, 1610, 1492, 1248, 1195. ¹H- and ¹³C-NMR (CDCl₃, 400, 100 MHz): Table 3. EI-MS: 342 (100, M⁺), 218 (5), 192 (16), 135 (100), 123 (66). HR-EI-MS: 342.1110 (calc. for C₁₉H₁₈O₆, 342.1103).

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